

# Characterization of Two Novel High Capacity, High Resolution Strong Ion Exchange Resins

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### Introduction

- TOYOPEARL GigaCap® Q-650S media is a high capacity, strong quaternary ammonium anion exchange resin optimized for the purification and polishing of both small and large proteins.
- TOYOPEARL GigaCap S-650S media is a high capacity, strong sulfonic cation exchange resin optimized specifically for the purification and polishing of IgG and smaller proteins.
- TOYOPEARL® HW-65 resin, the polymeric base bead for both resins, is chemically modified to provide higher capacity. Both resins have a pressure rating of 0.3 MPa, and they are stable in the pH range 3-13.
- Comparisons were made between the TOYOPEARL GigaCap Q-650S and the TOYOPEARL GigaCap S-650S to their M-grade counterparts showing increased selectivity as well as to other commercially available high resolution resins.

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## **Anion Exchange Resin Binding Capacity Comparisons**

Resin	Particle Size (µm)	pH Stability	Base Bead	Ion Exchange Capacity (meq/L)	Binding Capacity (g/L)		DBC Recovery	DBC Elution Volume
					Static	Dynamic <sup>1</sup>	(%)	(CV)
TOYOPEARL GigaCap Q-650S	20 - 50	3 - 13	Polymethacrylic	0.20	200	191	99	1.7
TOYOPEARL GigaCap Q-650M	50 - 100	3 - 13	Polymethacrylic	0.17	191	172	97	15.8
Capto™ Q ImpRes	36 - 44	2 - 12	Agarose	0.12	92	40	100	ND <sup>2</sup>
Q Sepharose™ HP	24 - 44	2 - 12	Agarose	0.15	114	81	99	ND <sup>2</sup>

<sup>1</sup>Dynamic binding capacities were determined at 10% breakthrough <sup>2</sup>Values not determined

### **Dynamic Binding Capacity (DBC) Conditions:**

Column size:  $6 \text{ mm ID} \times 4 \text{ cm}$ 

Mobile phase A: 50 mmol/L Tris-HCl buffer, pH 8.5

B: mobile phase A + 0.5 mol/L NaCl

Linear velocity: 212 cm/hr
Detection: UV @ 280 nm
Sample: 1.0 g/L BSA

### Static Binding Capacity (SBC) Conditions:

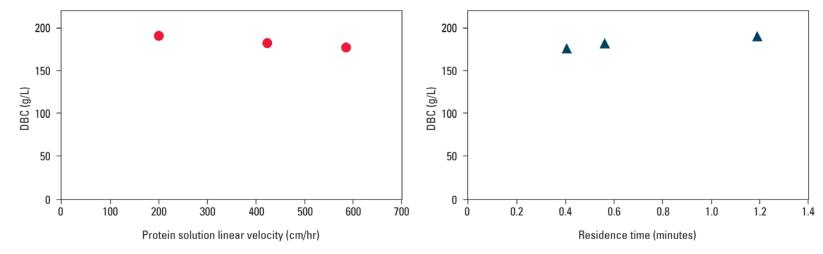
Adsorption buffer: 50 mmol/L Tris-HCl, pH 8.5

Protein concentration: 10.0 g/L

- The TOYOPEARL GigaCap series has a significantly higher binding capacity compared to GE Healthcare's Capto Q ImpRes and Q Sepharose HP, while maintaining nearly 100% recovery.
- In addition, the TOYOPEARL GigaCap Q-650S elutes in a very narrow band compared to the TOYOPEARL GigaCap Q-650M, allowing more efficient handling in subsequent downstream processing steps.



## **DBC vs. Flow Rate: Residence Time**



### **DBC Conditions:**

Resin: TOYOPEARL GigaCap Q-650S

Column size: 6 mm ID × 4 cm

Mobile phase: A: 50 mmol/L Tris-HCl buffer, pH 8.5

B: mobile phase A + 0.5 mol/L NaCl

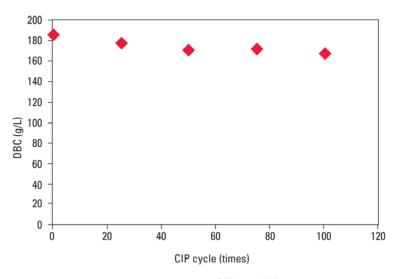
Linear velocity: 201, 424, or 585 cm/hr Residence time: 1.2 min, 0.6 min, or 0.4 min

Detection: UV @ 280 nm Sample: 1.0 g/L BSA

Efficient mass transfer kinetics allow the TOYOPEARL GigaCap Q-650S to maintain a high DBC even with increased linear velocities and decreased residence time.



## Stability of Resin After CIP



### **DBC** conditions:

Resin: TOYOPEARL GigaCap Q-650S

Column size:  $6 \text{ mm ID} \times 4 \text{ cm}$ 

Mobile phase: A: 50 mmol/L Tris-HCl, pH 8.5

B: mobile phase A + 1.0 mol/L NaCl

Flow rate: 212 cm/hr (1.0 mL/min)

Detection: UV @ 280 nm Sample: 1.0 g/L BSA

### **CIP** conditions:

Alkaline wash: 0.5 mol/L NaOH

Buffer wash: 50 mmol/L Tris + 0.5 mol/L NaCl, pH 8.5

Flow rate: 106 cm/hr (0.5 mL/min)

CIP wash volume: 27 CV/cycle

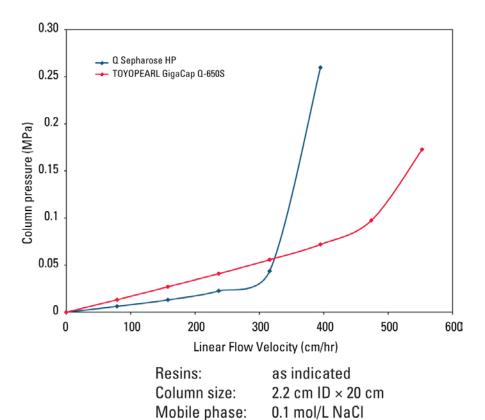
Contact time: 1 hr

Buffer wash volume: 10 CV/cycle

- The DBC of TOYOPEARL GigaCap Q-650S was measured after every set of 20 CIP cycles. Each CIP cycle consisted of a 27 CV (1 hour) wash with 0.5 mol/L NaOH followed by 10 CV of a pH 8.5 buffer.
- The DBC is maintained through at least 100 CIP cycles, showing excellent stability.



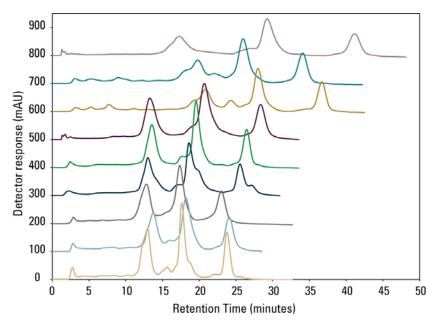
### **Pressure-Flow Curves**



- TOYOPEARL GigaCap Q-650S and Q Sepharose HP exhibited similar pressure-flow response for flow rates up to 300 cm/hr. Above 300 cm/hr, the Q Sepharose HP resin reached its critical velocity.<sup>1</sup> The particle sizes are 35 μm and 34 μm, respectively.
- The GigaCap Q-650S resin had a much higher critical velocity and can be operated at increased flow rates.



## **Selectivity Comparisons**





Column size:  $7.5 \text{ mm ID} \times 7.5 \text{ cm}$ 

Mobile phase: A: 50 mmol/L Tris-HCl, pH 8.5

B: mobile phase A + 1.0 mol/L NaCl

Gradient: 0-100% mobile phase B (120 min) Flow rate: 1.0 mL/min

Detection: UV @ 280 nm

Injection vol.: 100 μL

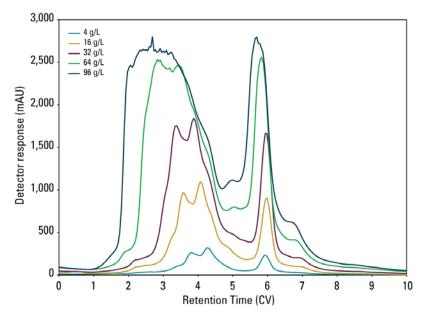
Sample: 2.9 g/L transferrin, 6.5 g/L ovalbumin,

10.0 g/L trypsin inhibitor

- The TOYOPEARL GigaCap Q-650S retains proteins to a higher conductivity than most of the resins tested.
- This data may indicate the TOYOPEARL GigaCap Q-650S resin is more salt tolerant than the other TOYOPEARL and TSKgel® Q-type resins.
- The retention is similar to Q Sepharose HP and Capto Q ImpRes. However, TOYOPEARL GigaCap Q-650S has improved selectivity compared to those two resins.



## **Standard Protein Loading**



Resin: TOYOPEARL GigaCap Q-650S

Column size:  $1.0 \text{ cm ID} \times 15.5 \text{ cm}$ 

Mobile phase: A: 50 mmol/L Tris-HCl, pH 8.7

B: mobile phase A + 0.5 mol/L NaCl

Gradient: 0-100% mobile phase B (10 CV)

Flow rate: 136 cm/hr (1.78 mL/min)

Detection: UV @ 280 nm Temperature: ambient Injection vol.: 0.4-9.6 CV

Sample: 4.5 g/L ovalbumin, 5.4 g/L trypsin inhibitor

Sample load: 4-96 g protein/L resin

Method: Column sanitized with 1.0 mol/L NaOH for

2.5 CV. Equilibrate column with mobile phase A

for 2 CV. Load proteins. Wash column with mobile phase A for 3 CV. Run 0-100%

mobile phase B gradient over 10 CV.

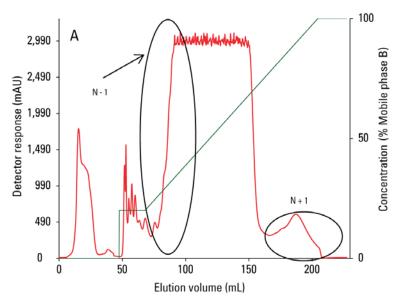
Clean column with 5 CV using 1.0 mol/L NaOH.

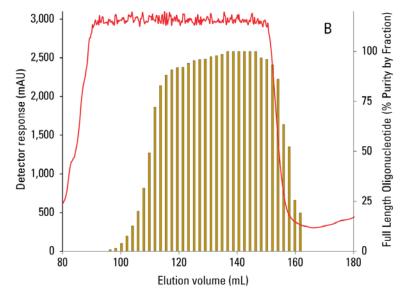
- Shown are chromatograms of TOYOPEARL GigaCap Q-650S resin with increasing protein load, ranging from 4 g protein/L resin to 96 g protein/L resin.
- At lower protein loads, the resin was able to separate various components in the ovalbumin.
- Even at 96 g/L protein loading, this resin has sufficient capacity and selectivity to separate the ovalbumin and its impurities from the trypsin inhibitor.

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## **Oligonucleotide Purification**





Resin: TOYOPEARL GigaCap Q-650S Column size:  $6.6 \text{ mm ID} \times 18 \text{ cm } (6.16 \text{ mL})$ 

Mobile phase: A: 20 mmol/L NaOH

B: mobile phase A + 3.0 mol/L NaCl

Gradient: step to 20% B (2 CV)

20% - 100% B (20 CV)

100% B (2 CV)

Flow rate: 200 cm/hr (1.14 mL/min)

Detection: UV @ 254 nm
Injection vol.: 4.63 mL (181.4 mg)
Sample: crude phosphorothioate deoxyribonucleotide

- TOYOPEARL GigaCap Q-650S effectively purified a crude phosphorothioate deoxyribonucleotide at 80% DBC loading.
- Although the UV trace at 254 nm went off scale, an enlarged image of the main oligonucleotide peak, when overlaid with a histogram showing HPLC results for fraction purity, shows the full length oligonucleotide completely contained in the main peak at 80% DBC loading.
- In addition, the TOYOPEARL GigaCap Q-650S is able to elute in a smaller pool volume compared to other resins (data not shown).



## Cation Exchange Resin Binding Capacity Comparisons

Resin	Particle Size (µm)	pH Stability	Base Bead	Ion Exchange Capacity (meq/L)	Binding Capacity (g/L)		DBC Recovery	DBC Elution Volume*
					Static	Dynamic*	(%)	(CV)
TOYOPEARL GigaCap S-650S	20 - 50	3 - 13	Polymethacrylic	0.24	177	164	99	4.0
TOYOPEARL GigaCap S-650M	50 - 100	3 - 13	Polymethacrylic	0.16	156	145	98	13.5
Capto SP ImpRes	36 - 44	2 - 12	Agarose	0.12	89	27	100	ND**
SP Sepharose HP	24 - 44	2 - 12	Agarose	0.15	105	65	100	ND**

<sup>\*</sup> Elution volumes calculated using lysozyme at 10% breakthrough.

#### **DBC Conditions:**

Column size: 6 mm ID × 4 cm

Mobile phase: A: 50 mmol/L acetate buffer, pH 4.7

B: mobile phase A + 0.5 mol/L NaCl

Linear velocity: 212 cm/hr
Detection: UV @ 280 nm
Sample: 1.0 g/L  $\gamma$ -globulin

### **SBC Conditions:**

Adsorption buffer: 50 mmol/L acetate buffer, pH 4.7

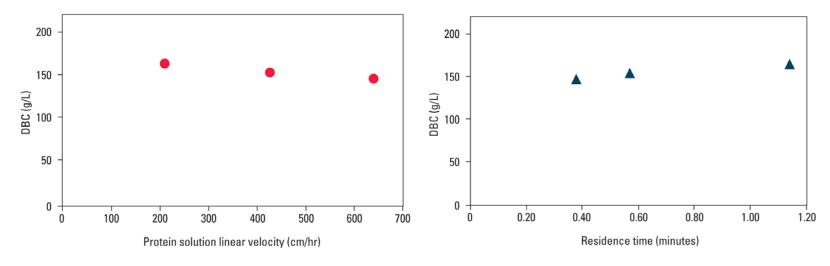
Sample:  $10.0 \text{ g/L } \gamma$ -globulin

- Both TOYOPEARL GigaCap resins have a significantly higher binding capacity compared to GE Healthcare's Capto SP ImpRes and SP Sepharose HP while maintaining nearly 100% recovery.
- In addition, the TOYOPEARL GigaCap S-650S elutes in a very narrow band compared to TOYOPEARL GigaCap S-650M, allowing for more efficient handling in subsequent downstream processing steps.

<sup>\*\*</sup> Values not determined.



## **DBC vs. Flow Rate: Residence Time**



### **DBC Conditions:**

Resin: TOYOPEARL GigaCap S-650S

Column size: 6 mm ID × 4 cm

Mobile phase: A: 50 mmol/L acetate pH 4.7

B: mobile phase A + 0.5 mol/L NaCl

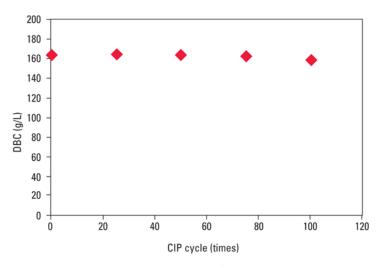
Linear velocity: 211, 425, or 637 cm/hr Residence time: 1.1 min, 0.6 min, or 0.4 min

 $\begin{array}{lll} \mbox{Detection:} & \mbox{UV @ 280 nm} \\ \mbox{Sample:} & \mbox{1.0 g/L } \gamma\mbox{-globulin} \end{array}$ 

Efficient mass transfer kinetics allow the TOYOPEARL GigaCap S-650S to maintain a high DBC even with increased linear velocities and decreased residence time.



## Stability of Resin After CIP



#### **DBC** conditions:

Resin: TOYOPEARL GigaCap S-650S

Column size:  $6 \text{ mm ID} \times 4 \text{ cm}$ 

Mobile phase: A: 50 mmol/L acetate, pH 4.7

B: mobile phase A + 1.0 mol/L NaCl

Flow rate: 212 cm/hr (1.0 mL/min)

Detection: UV @ 280 nm Sample: 1.0 g/L γ-globulin

### **CIP** conditions:

Alkaline wash: 0.5 mol/L NaOH

Buffer wash: 50 mmol/L acetate + 0.5 mol/L NaCl, pH 4.7

Flow rate: 106 cm/hr (0.5 mL/min)

CIP wash volume: 27 CV/cycle

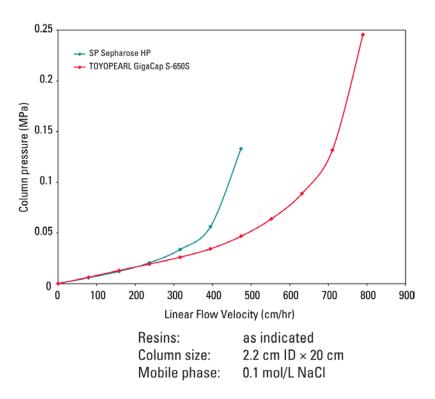
Contact time: 1 hr

Buffer wash volume: 10 CV/cycle

- The DBC of TOYOPEARL GigaCap S-650S was measured after every set of 20 CIP cycles.
- Each CIP cycle consisted of a 27 CV (1 hour) wash with 0.5 mol/L NaOH followed by 10 CV of a pH 8.5 buffer.
- The DBC is maintained through at least 100 CIP cycles, showing excellent stability.



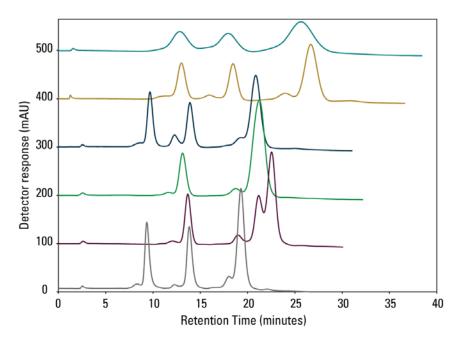
### **Pressure-Flow Curves**



- TOYOPEARL GigaCap S-650S and SP Sepharose HP exhibited similar pressure-flow responses for flow rates up to 225 cm/hr. Above 450 cm/hr, the SP Sepharose HP resin reached its critical velocity. The particle sizes are 35 μm and 34 μm, respectively.
- The GigaCap S-650S resin had a much higher critical velocity and can be operated at increased flow rates.



## **Selectivity Comparisons**



— TOYOPEARL GigaCap S-650M — TOYOPEARL GigaCap S-650S — TOYOPEARL SP-650S — Capto SP ImpRes — SP Sepharose HP — TSKgel SP-5PW (20)

Column size:  $7.5 \text{ mm ID} \times 7.5 \text{ cm}$ 

Mobile phase: A: 20 mmol/L phosphate, pH 7.0

B: mobile phase A + 1.0 mol/L NaCl

Gradient: 0-100% mobile phase B (60 min)

Flow rate: 1.0 mL/min Detection: UV @ 280 nm

Injection vol.: 20 µL

Samples: ribonuclease A, 9.8 g/L

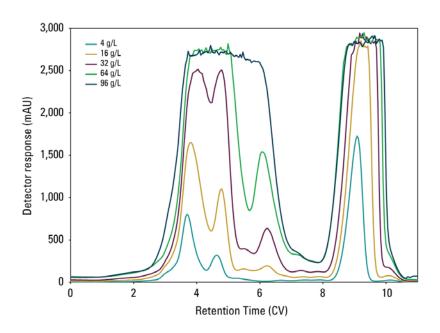
cytochrome C, 3.6 g/L

lysozyme, 6.4 g/L

- The TOYOPEARL GigaCap S-650 resins retain proteins at higher conductivities than the TSKgel SP-5PW (20) and TOYOPEARL SP-650S.
- Although the TOYOPEARL GigaCap S-650S shows similar retentivity to Capto SP ImpRes, SP Sepharose HP, and TOYOPEARL GigaCap S-650M, the TOYOPEARL GigaCap S-650S elutes with improved selectivity and lower pool volumes.



## **Standard Protein Loading**



Resin: TOYOPEARL GigaCap S-650S

Column size:  $1.0 \text{ cm ID} \times 15.8 \text{ cm}$ 

Mobile phase: A: 20 mmol/L sodium phosphate, pH 4.7

B: mobile phase A + 0.5 mol/L NaCl

Gradient: 0-100% mobile phase B (10 CV)

Flow rate: 300 cm/hr
Detection: UV @ 280 nm
Temperature: ambient
Injection vol.: 0.4-9.6 CV

Sample:  $\alpha$ -chymotrypsin, 2.0 g/L

lysozyme, 2.0 g/L

Sample load: 4-96 g protein/L resin

Method: Column sanitized with 1.0 mol/L NaOH for

2.5 CV. Equilibrate column with mobile phase A for 2 CV. Load proteins. Wash column with mobile phase A for 3 CV. Run 0-100% mobile phase B gradient over 10 CV. Clean column

with 5 CV using 1.0 mol/L NaOH.

- Shown are chromatograms of TOYOPEARL GigaCap Q-650S resin with increasing protein load, ranging from 4 g protein/L resin to 96 g protein/L resin.
- At lower protein loads, the resin was able to separate various components in the α-chymotrypsin.
- Even at 96 g/L loading, the resin still had sufficient capacity and selectivity to separate the α-chymotrypsin and its impurities from lysozyme.



### **Conclusions**

- TOYOPEARL GigaCap Q-650S and TOYOPEARL GigaCap S-650S are high capacity and high resolution strong ion exchange resins useful in both purification and polishing steps.
- Like their M-grade counterparts, the TOYOPEARL GigaCap Q-650S and TOYOPEARL GigaCap S-650S resins have high binding capacities and improved elution kinetics, resulting in a decreased elution pool volume.
- Dynamic binding capacity of the TOYOPEARL GigaCap Q-650S was higher than the TOYOPEARL GigaCap Q-650M at around 191 g BSA/L resin, compared to 172 g BSA/L resin.
- Dynamic binding capacity of the TOYOPEARL GigaCap S-650S resin was also higher than its M-grade counterpart, coming in at around 164 g γ-globulin/L resin, compared to 145 g γglobulin/L resin.
- The elution kinetics for the TOYOPEARL GigaCap Q-650S resin are significantly improved over the TOYOPEARL GigaCap Q-650M. The BSA eluted in approximately 1.7 CV, almost an order of magnitude less than the TOYOPEARL GigaCap Q-650M, which eluted in 15.8 CV.
- The TOYOPEARL GigaCap S-650S eluted lysozyme in approximately 4.0 CV, whereas the TOYOPEARL GigaCap S-650M eluted lysozyme in 13.5 CV.
- Both resins have a high capacity, even at increased flow rates.
- Both resins also exhibit reasonable selectivity, allowing for some separation of the test proteins, even at a protein load of 96 g total protein/L resin.



### Reference

<sup>1</sup> Stickel JJ; Fotopoulos A. Pressure-flow relationships for packed beds of compressible chromatography media at laboratory and production scale. *Biotechnol Prog* **2001**, *17*, 744-751.